

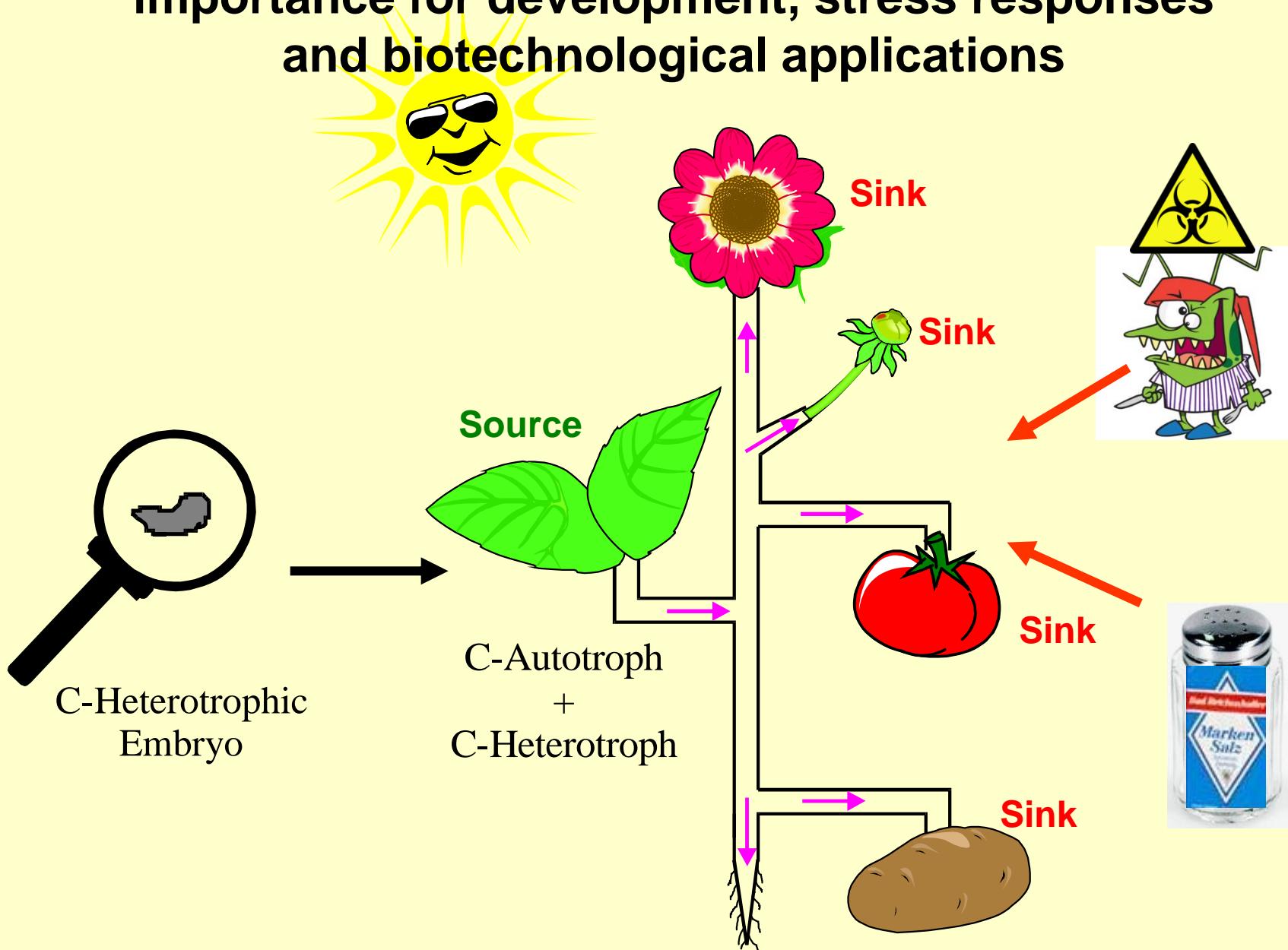


Assimilate partitioning in higher plants and Plant Phenomics

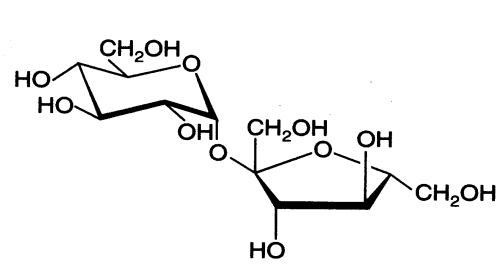
Eric van der Graaff and Thomas Roitsch
University of Copenhagen
Department of Plant and Environmental Sciences



Assimilate partitioning in higher plants: importance for development, stress responses and biotechnological applications

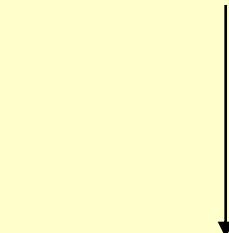


The Invertase Isoenzymes



Sucrose

α -D-Glucopyranosyl- β -D-fructofuranoside



Glucose + Fructose

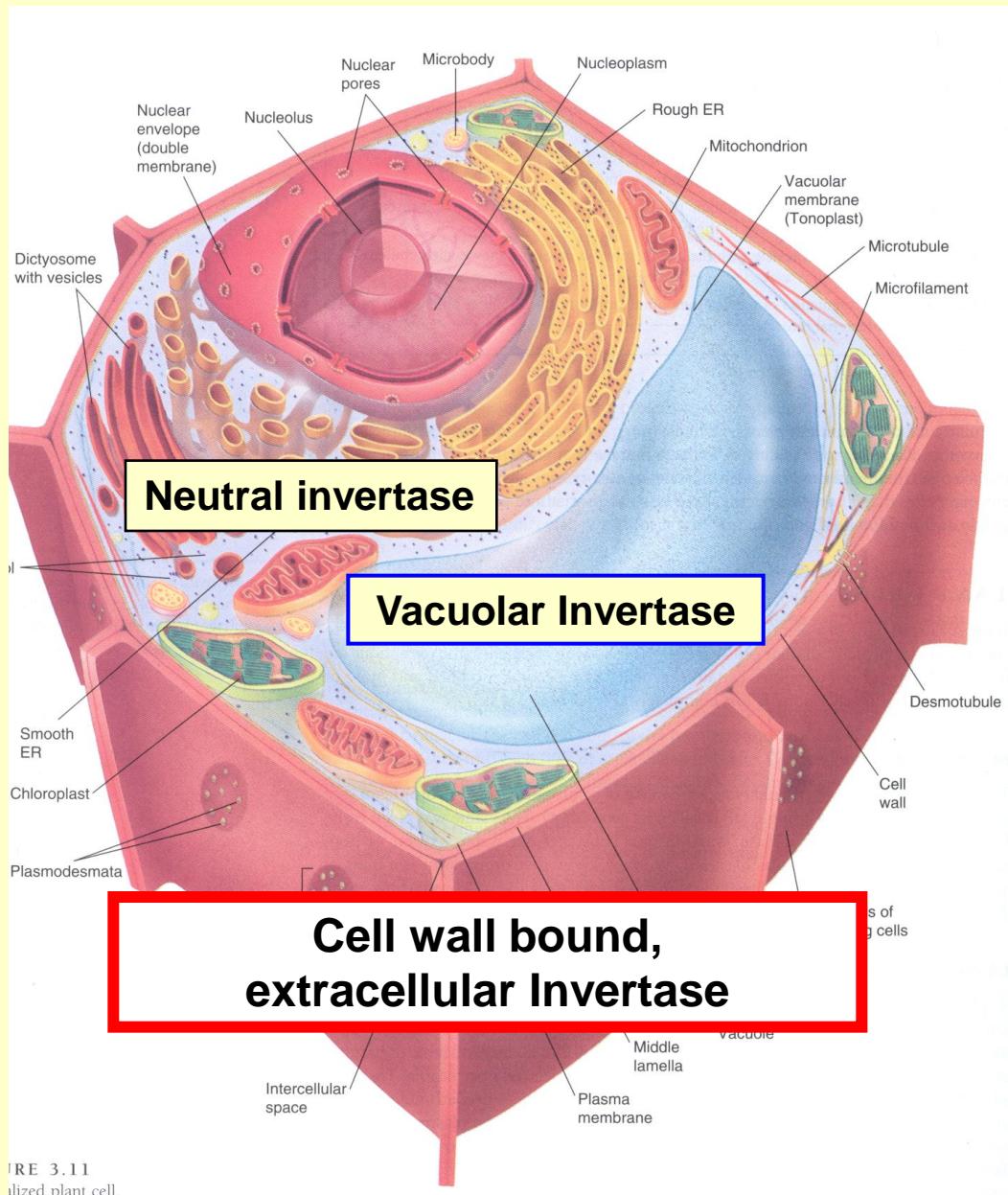
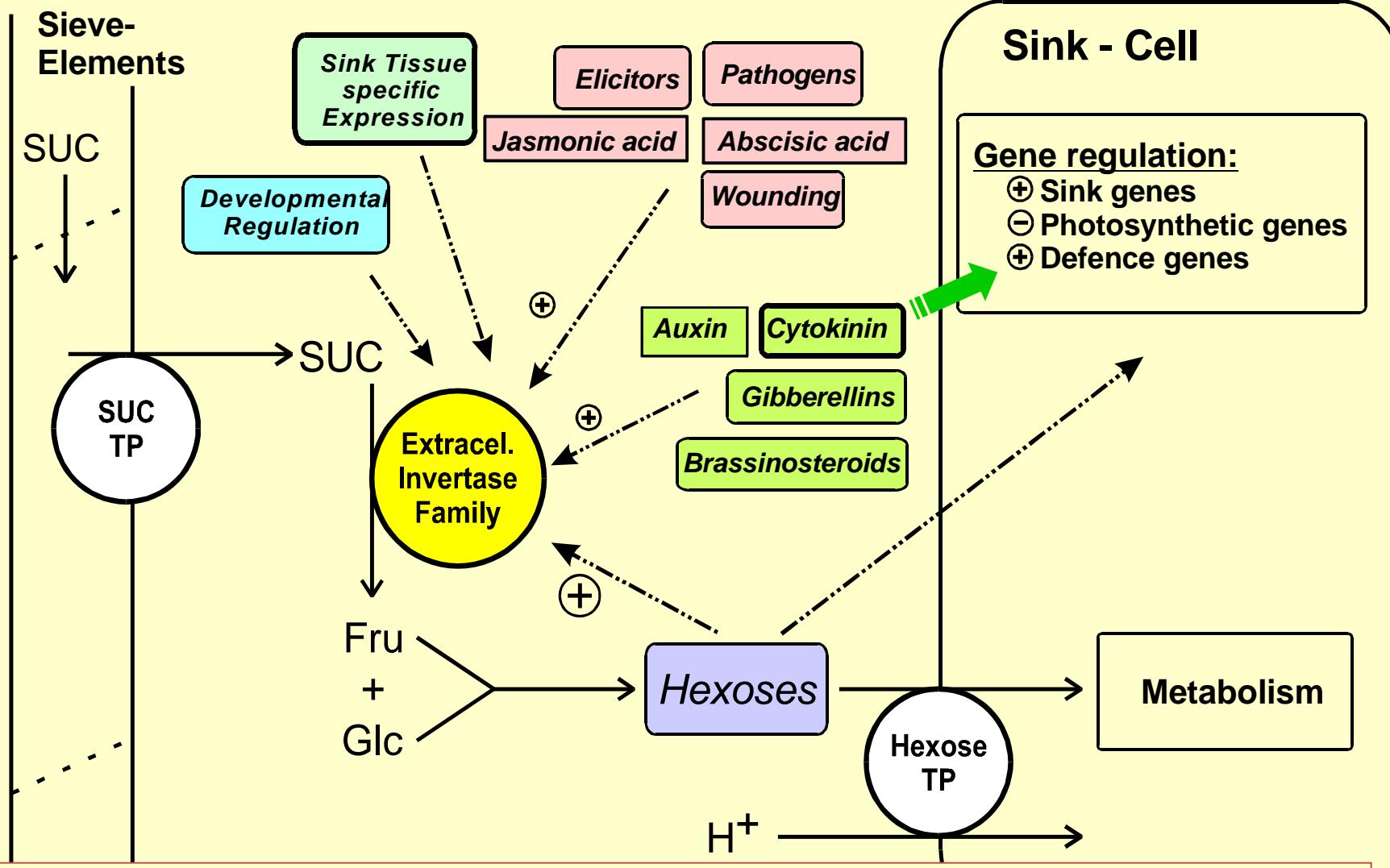


FIGURE 3.11
A generalized plant cell.

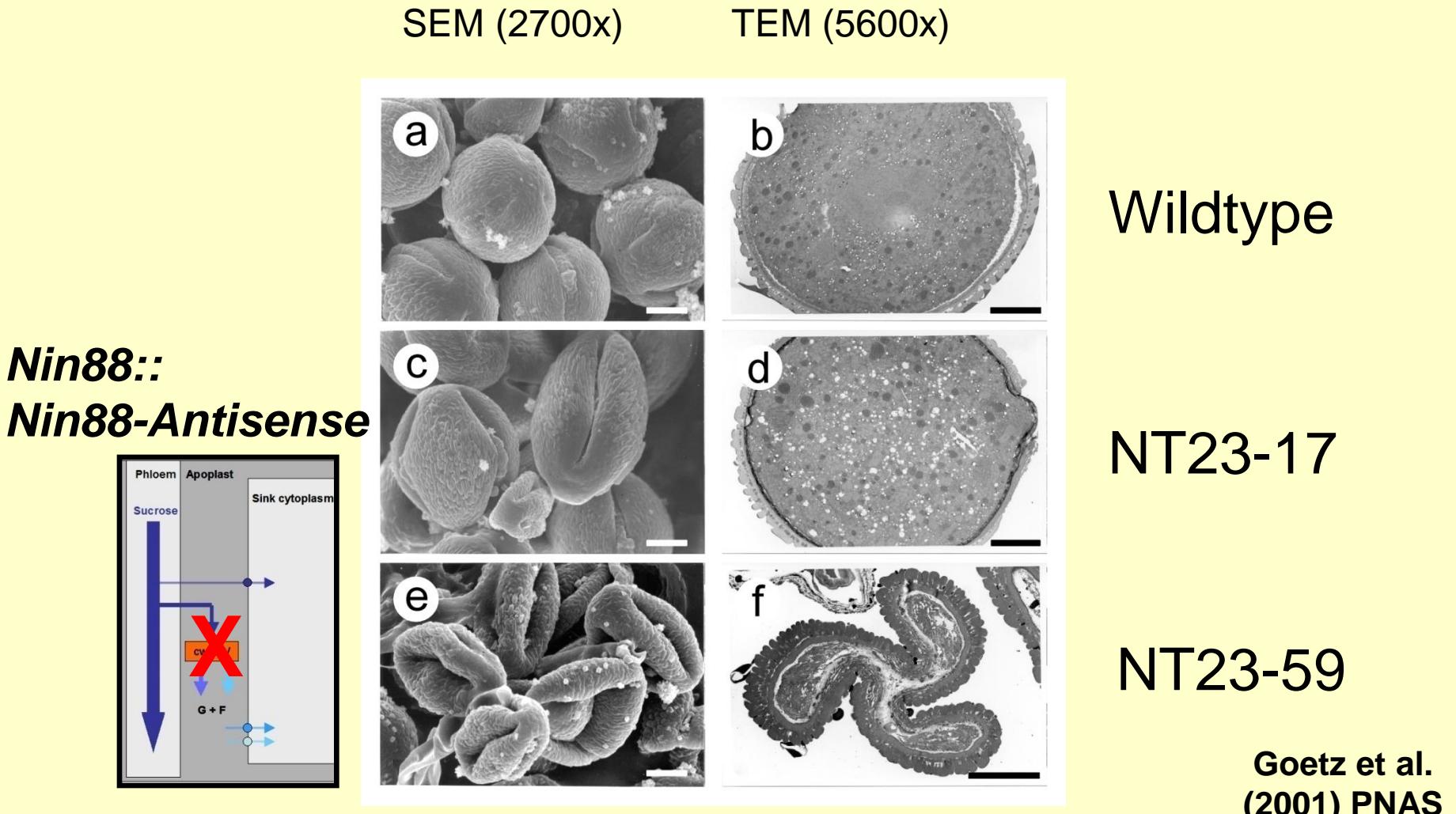
Extracellular Invertase: Key metabolic Enzyme and PR Protein



Roitsch (1999) COPS 2, 98f; Roitsch et al. (2004) TIPS 9, 607f; Berger et al. (2007) JXB 58, 4019f
Albacete et al. (2011) Phyton 50, 181; Großkinsky et al. (2012) Plant Sci. 195, 54f

Extracellular invertase is essential for pollen development

Antisense-Repression of Extracellular Invertase *Nin88*
results in an Arrest of Pollen Development and male sterile Plants



Relation between phytohormone action and carbohydrate metabolism

Extracellular Invertase is essential for the delay of senescence by cytokinins

Cytokinin → Extracellular Invertase → Delay of Senescence

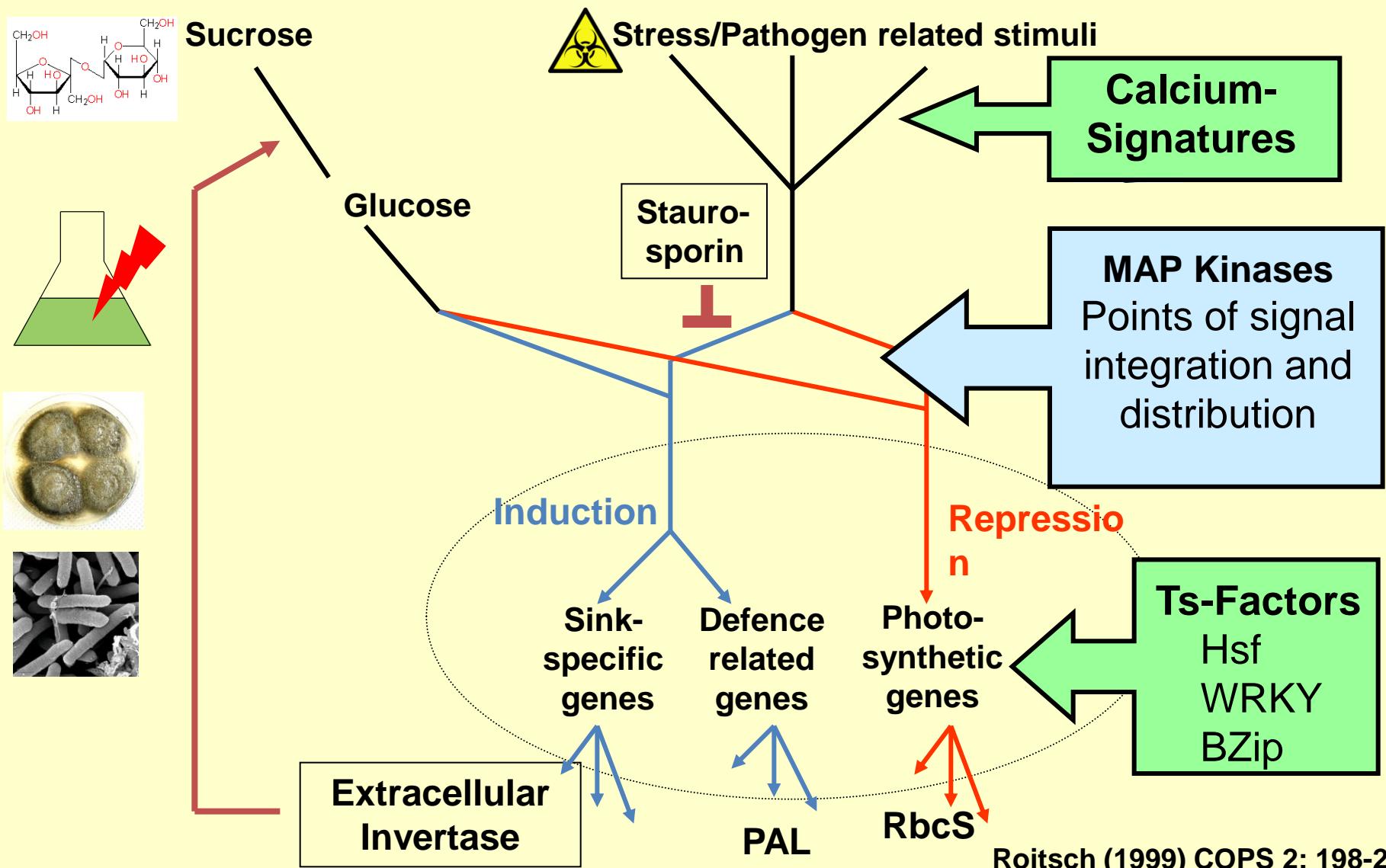
SAG12:
Cin1

W 38



Balibrea et al..
(2004) Plant Cell

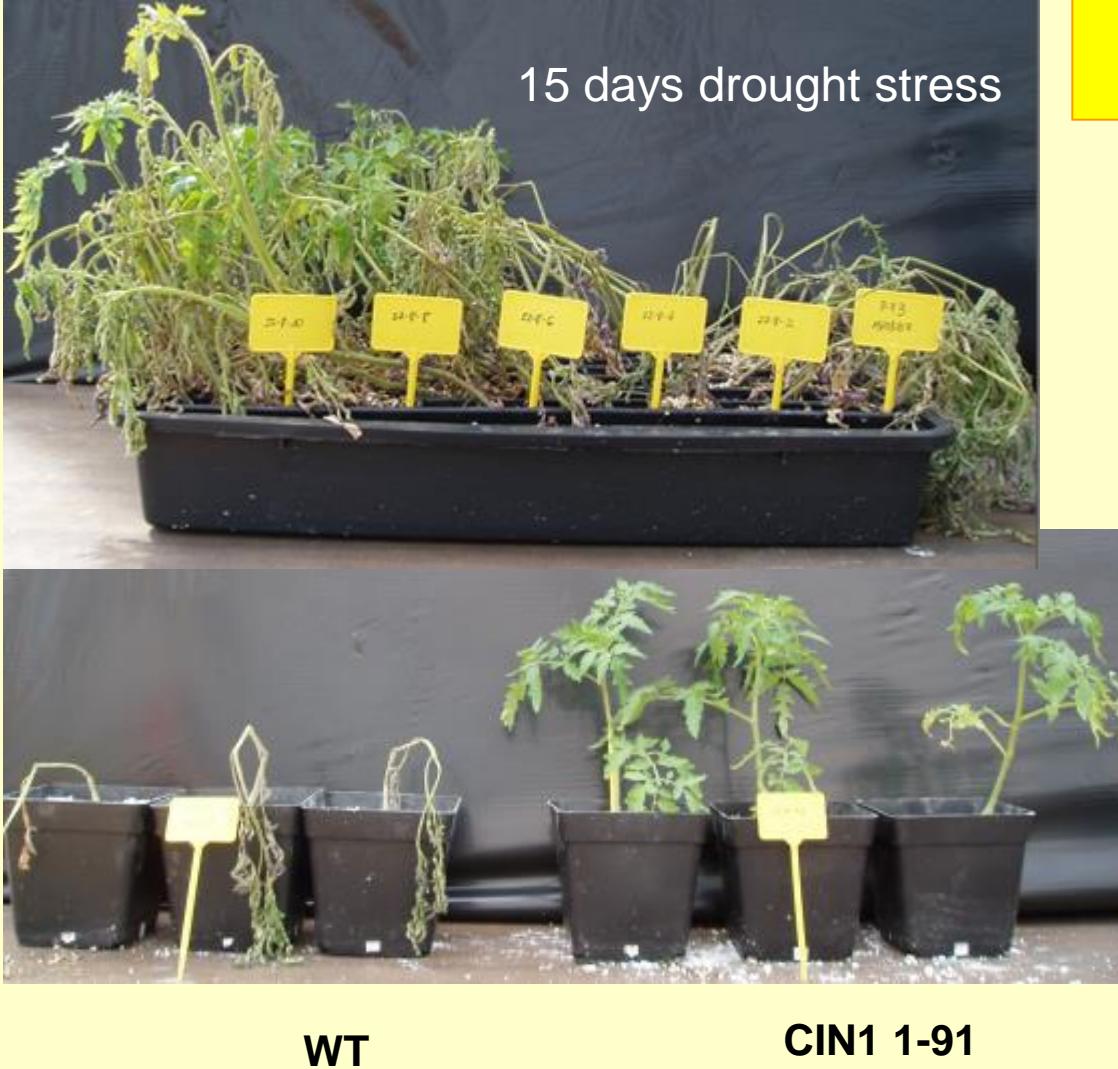
Co-ordinated regulation of source/sink relations and defence response by sugars and stress related stimuli



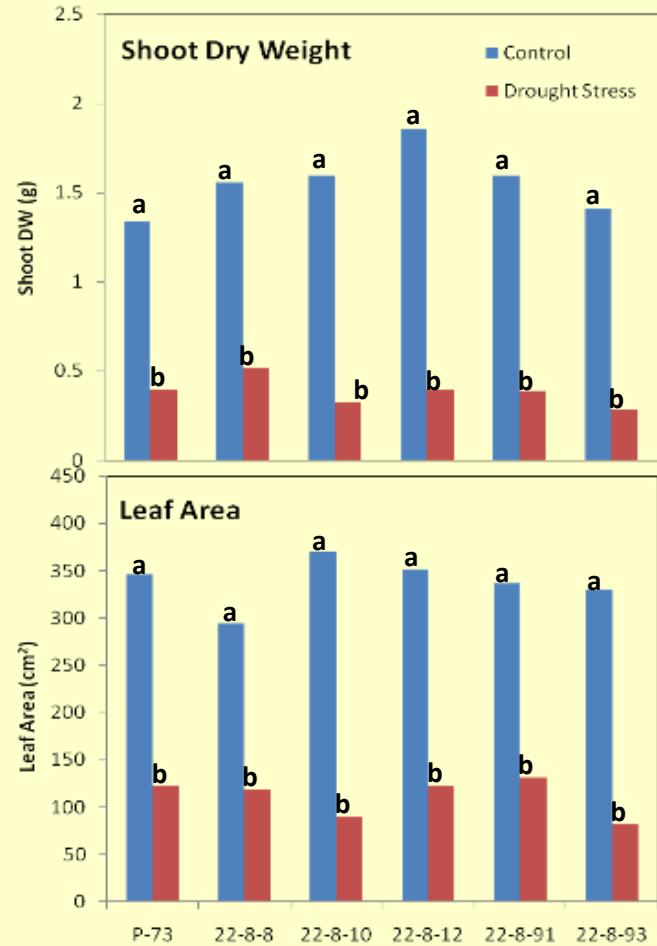
Roitsch (1999) COPS 2: 198-206

Ehness et al. (1997) Plant Cell 9: 1825

Ectopic expression of Extracellular invertase results in extreme drought stress tolerance



Drought stress tolerance in *C/N1* is not accompanied by a yield penalty under control conditions



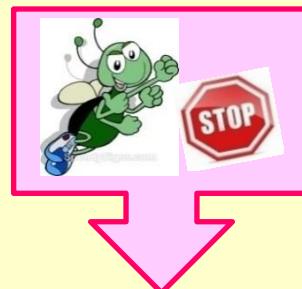
Cytokinins stimulate immunity via stimulation of phytoalexin synthesis within the Phytohormones Networks

Phytohormone defence backbone:

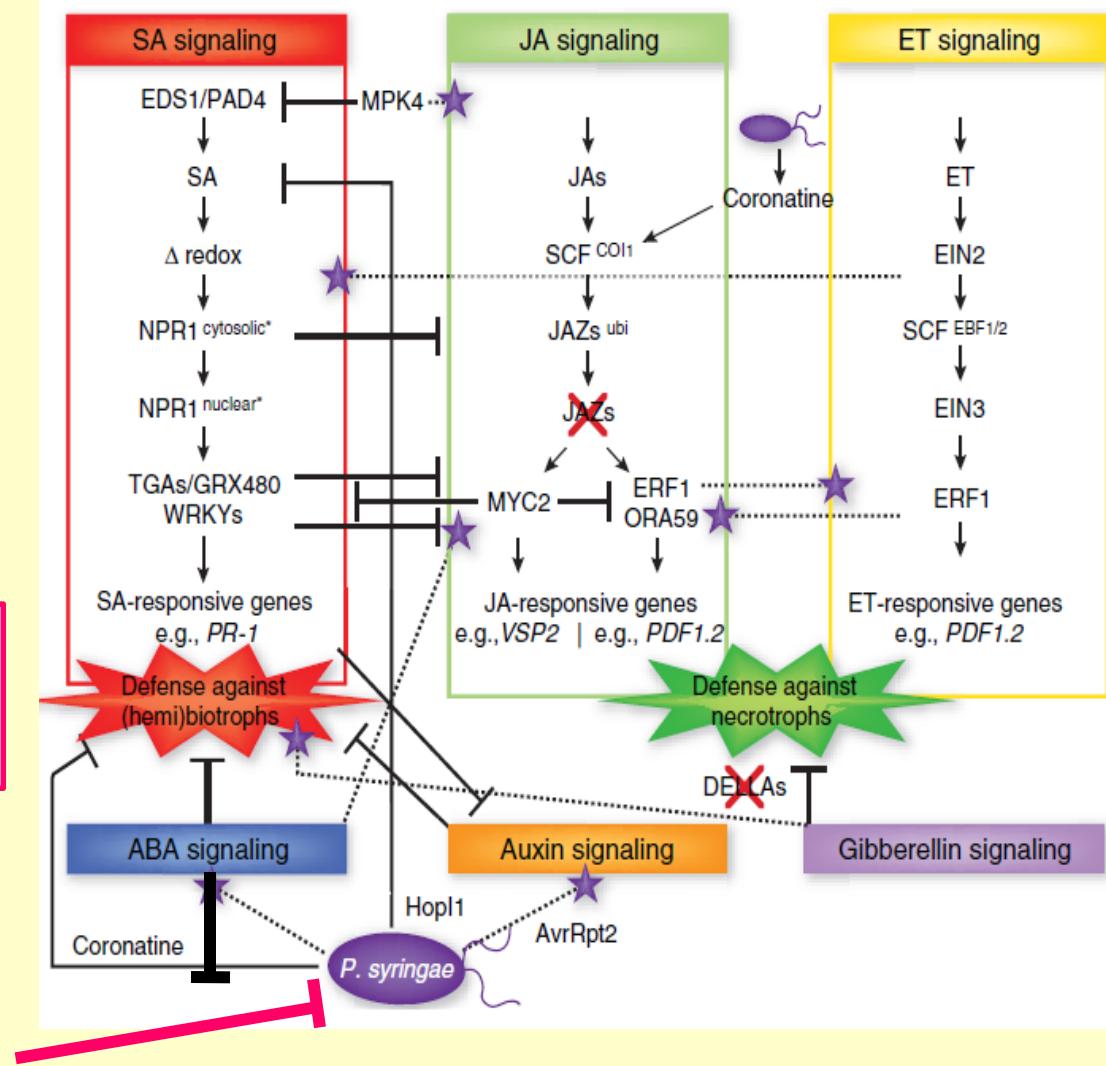
- Salicylic Acid
- Jasmonic Acid
- Ethylene

Additional interactions :

- Abscisic Acid
 - Auxin
- Gibberellin

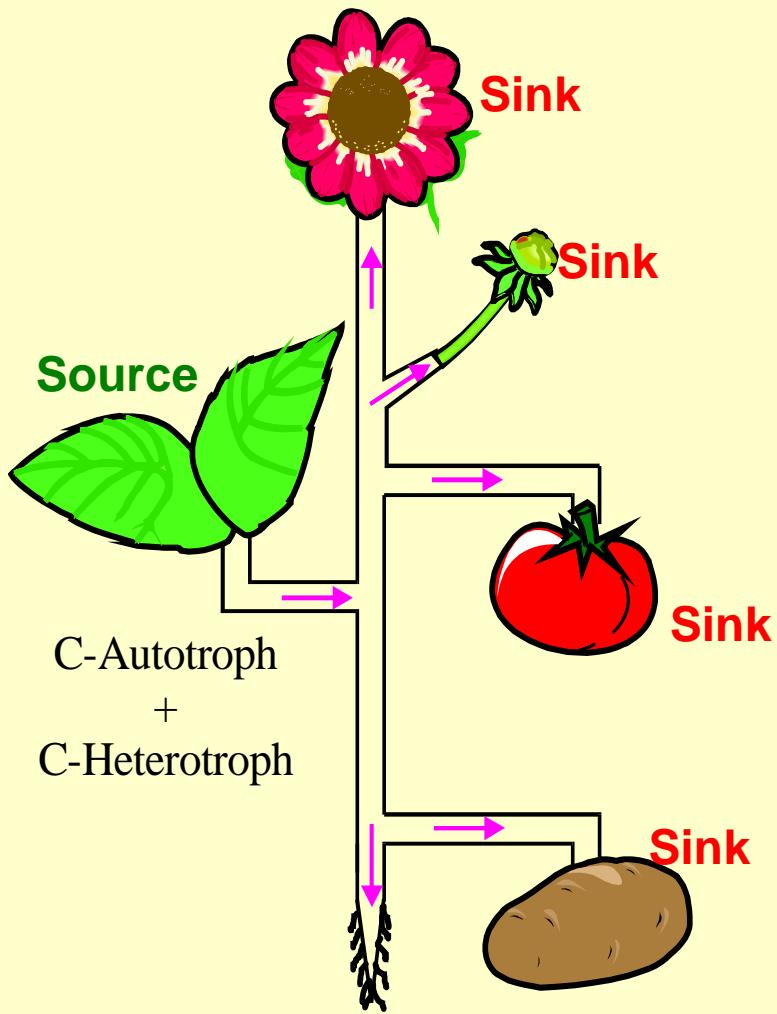


+ Cytokinin signalling
Phytoalexins

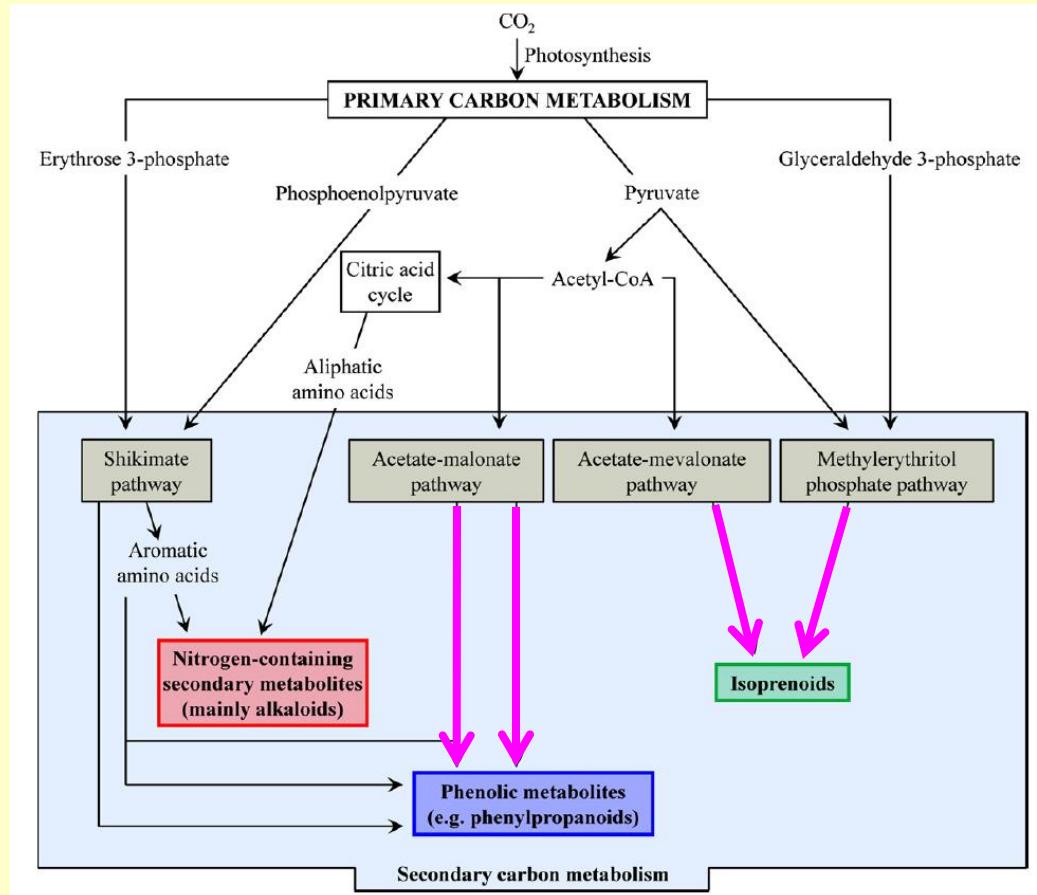


Different scales of carbohydrate partitioning

(A) Long distance transport from source and sink tissues



(B) Metabolic channeling from primary to secondary metabolism

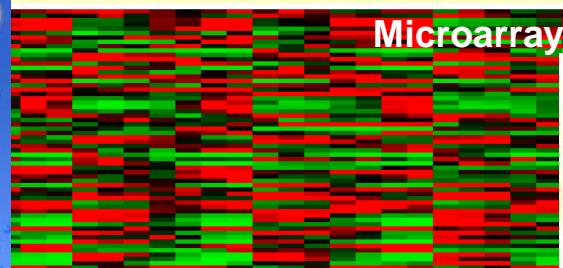
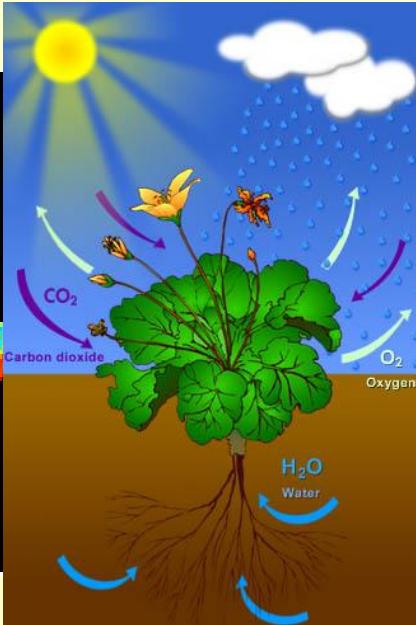
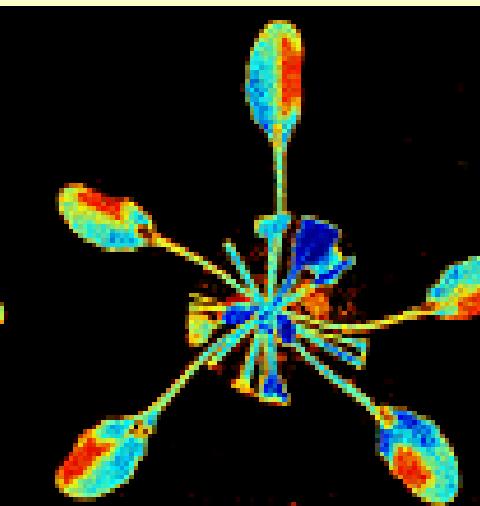




Phenomics

Development and application of high-throughput, multi-dimensional and dynamic phenotyping

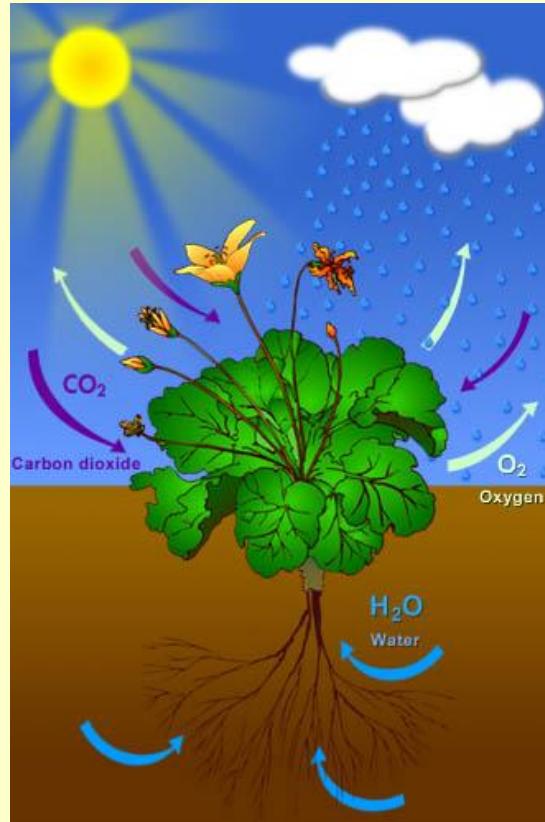
- Gene to phenotype modelling -



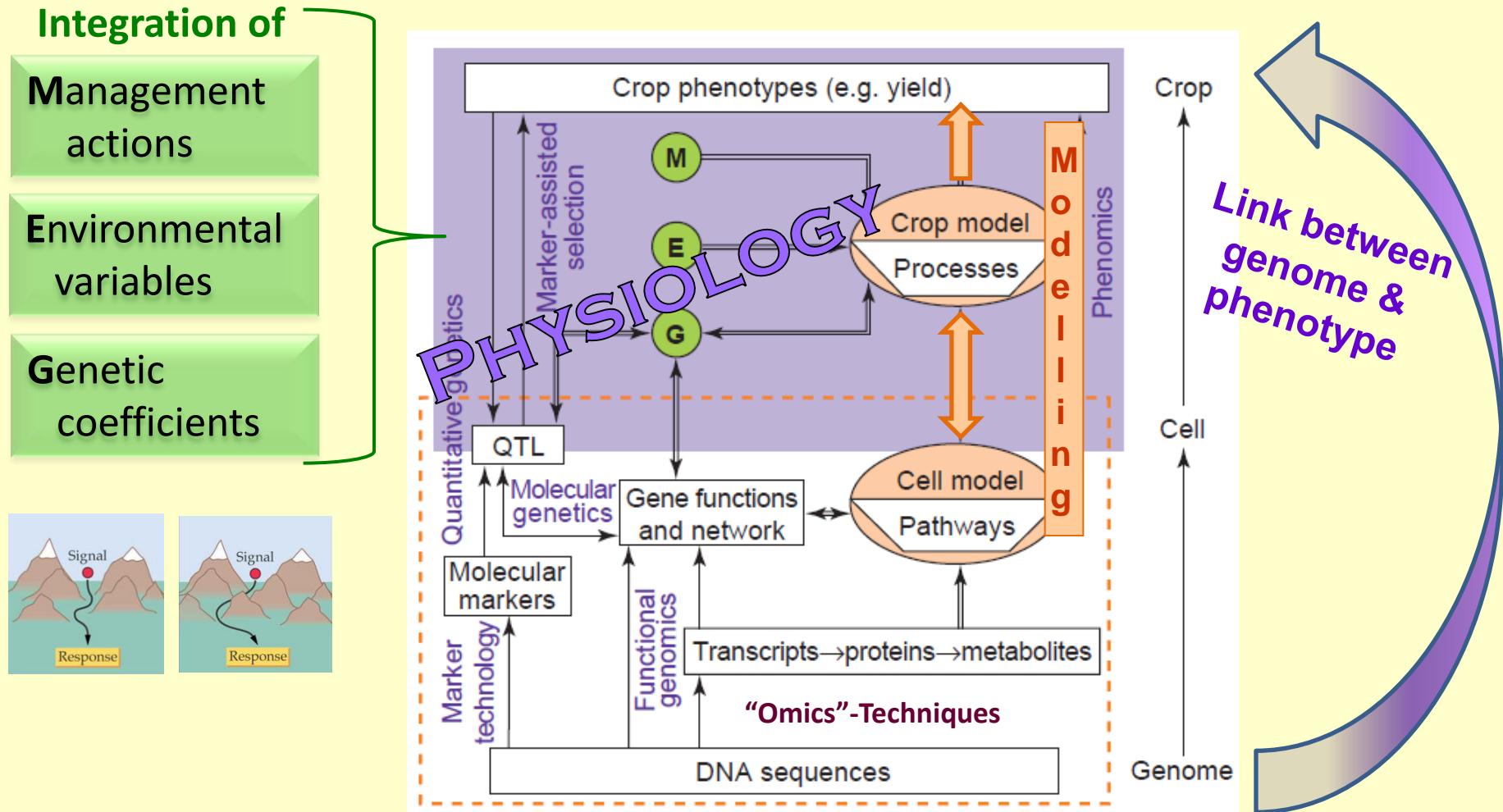
semi-qRT-PCR

Phenotypic variation

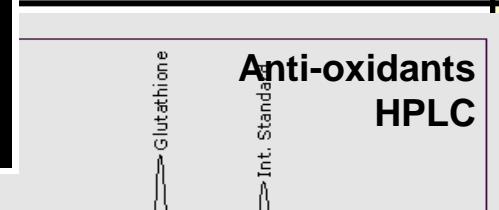
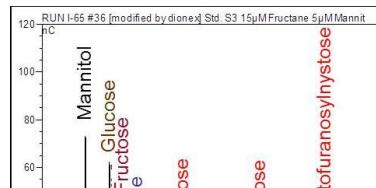
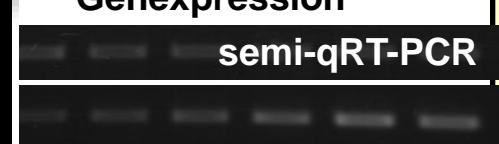
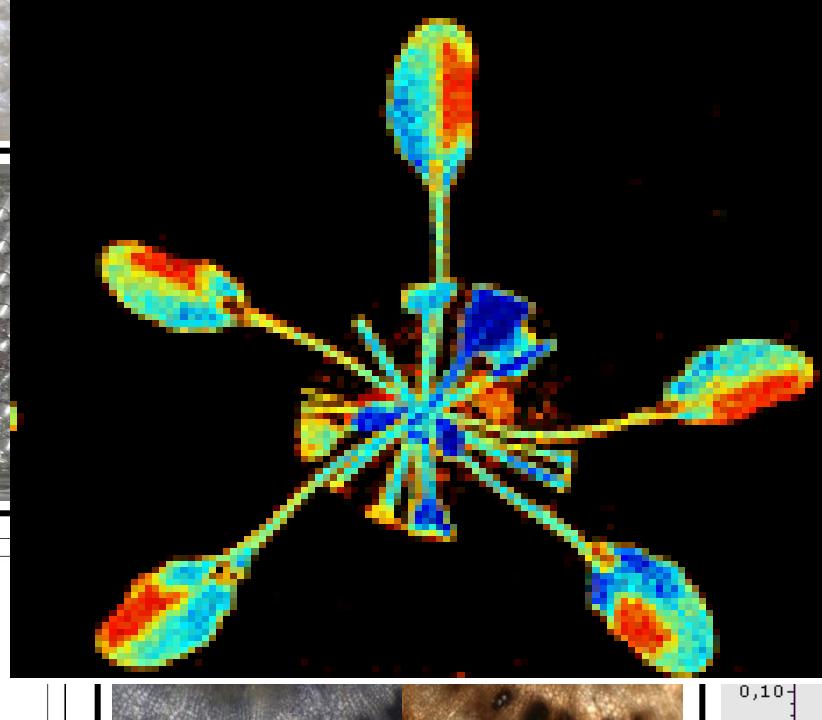
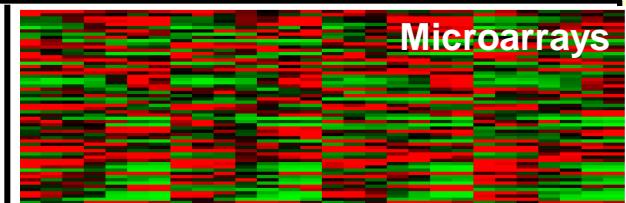
... is produced through a complex web of interactions
between
Genotypes x Environment x Management



Plant Physiology – Key Interface between genome and quantitative traits that determines harvest yield and quality



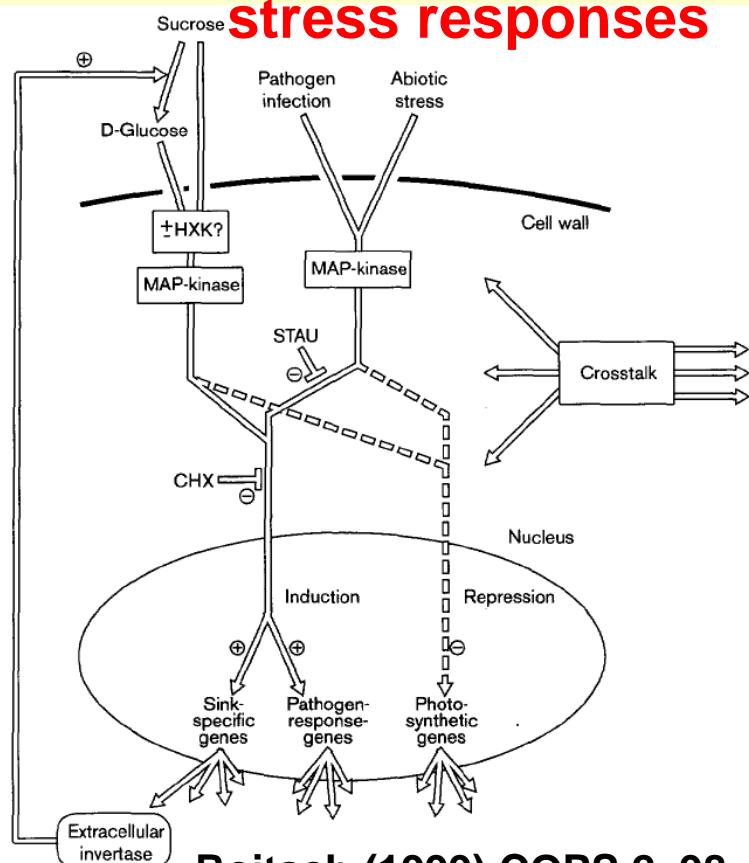
Integration of non-invasive signatures with physiological phenotyping: „Plant physiology meets biophysics“



Linking hyper-scale, non-invasive imaging to temporal and spatial dynamics of physiological processes and DNA-based markers

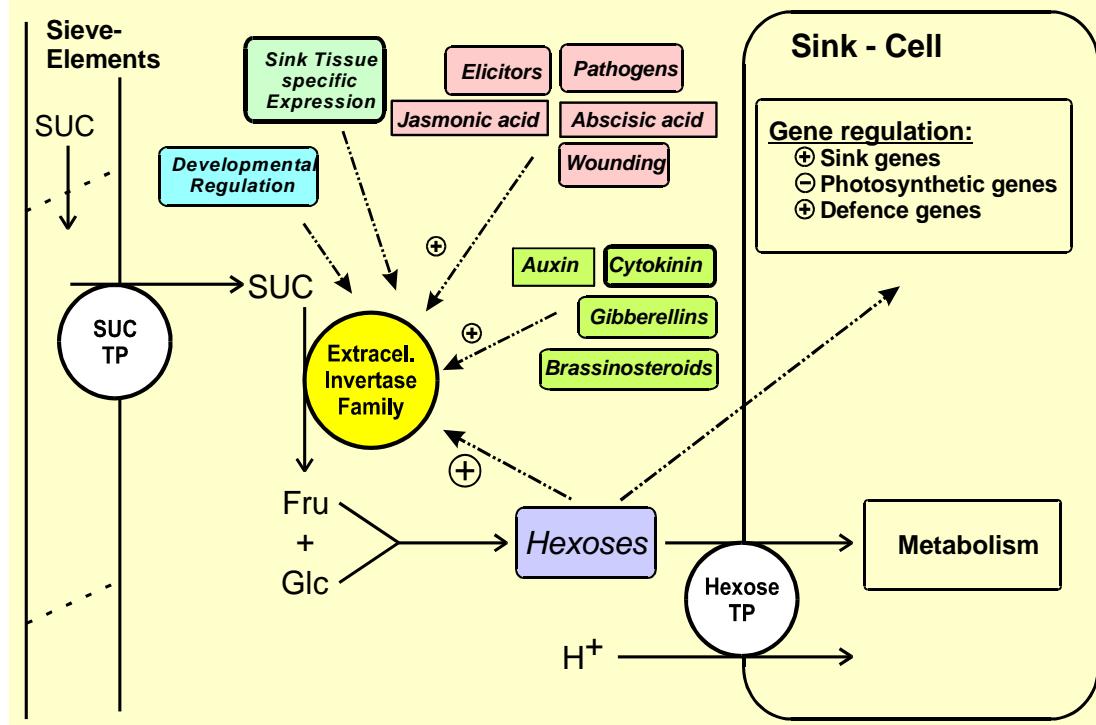
Development & stress responses are linked to plant carbohydrate metabolism

Co-ordinated regulation of source-sink relations & stress responses



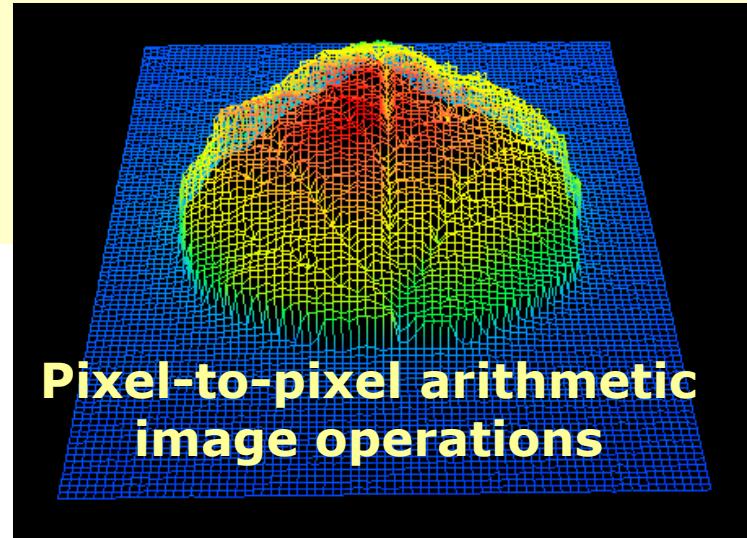
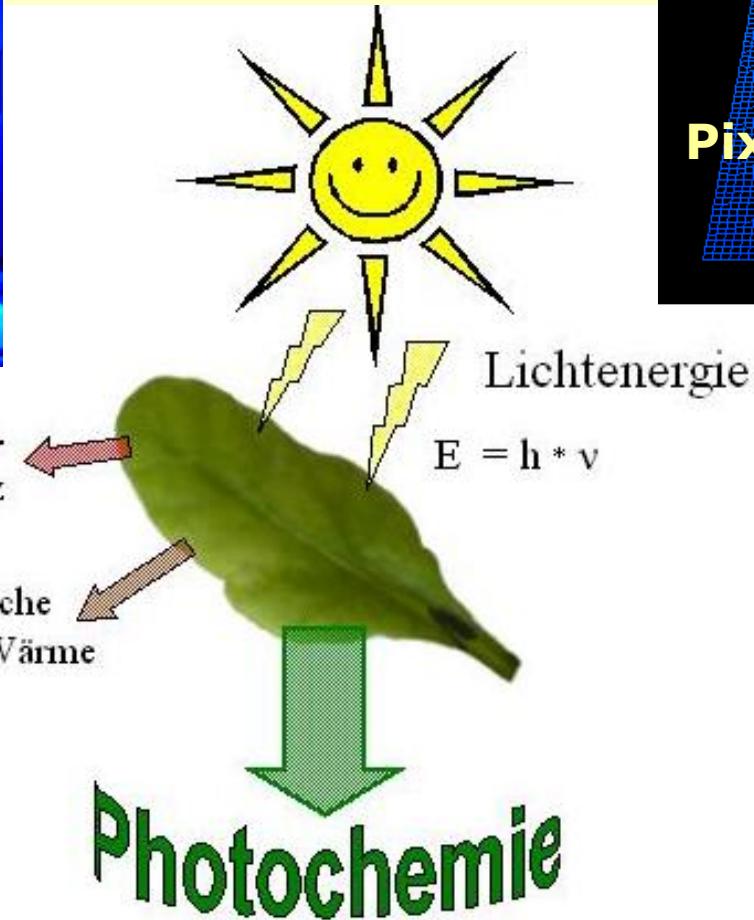
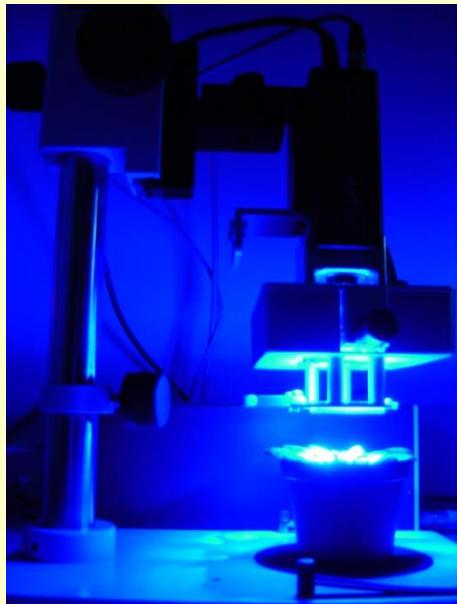
Roitsch (1999) COPS 2, 98
Berger et al. (2007) JXB 58, 4019
Albacete et al. (2011) Phyton 50,181

Extracellular Invertase: key metabolic enzyme and stress response protein



Roitsch et al. (1995) PlantPhys 108: 285
Roitsch et al. (2004) TIPS 9, 607
Albacete et al. (2011) Phyton 50,181

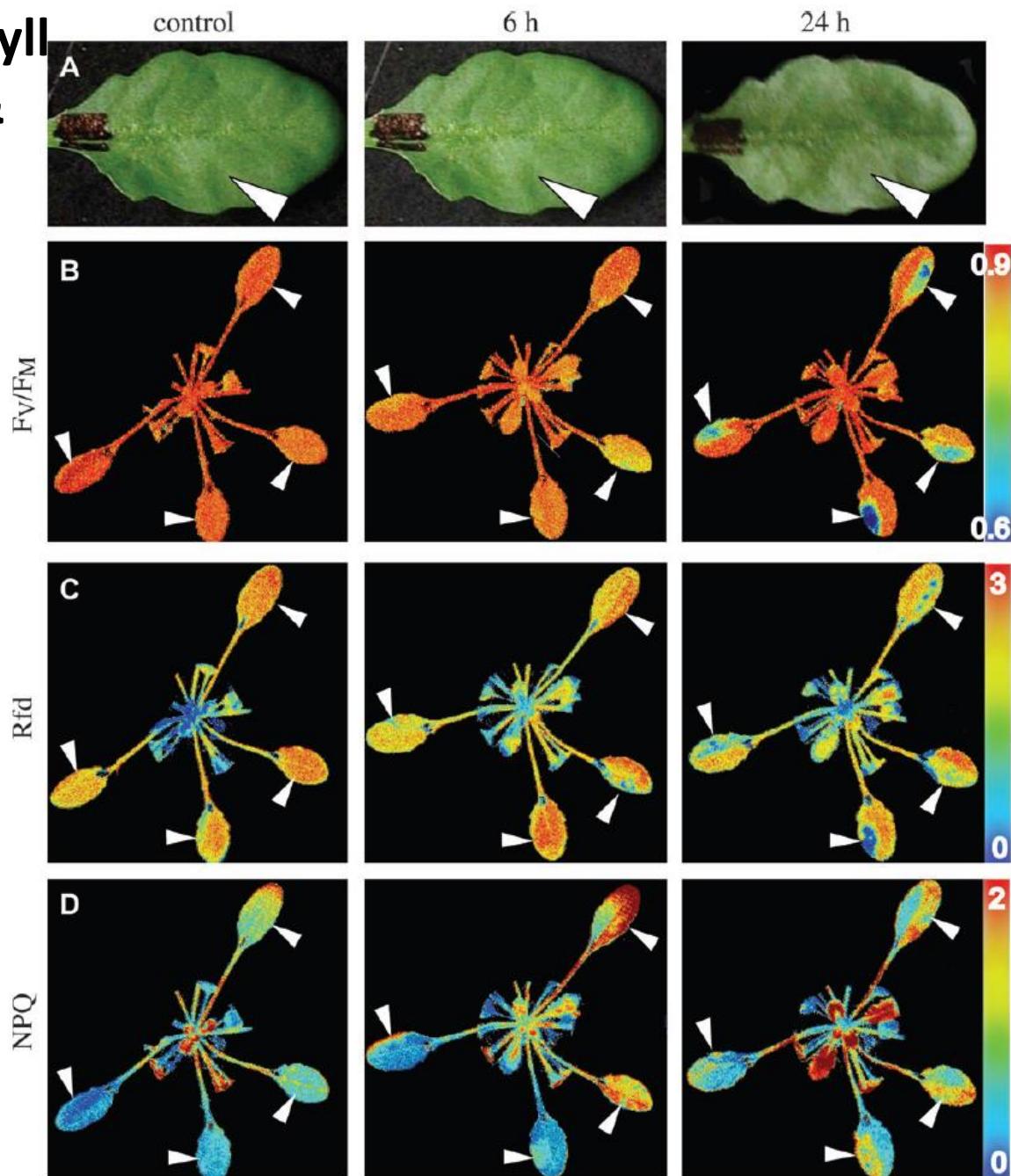
Chlorophyll Fluorescence Imaging



Advantages:

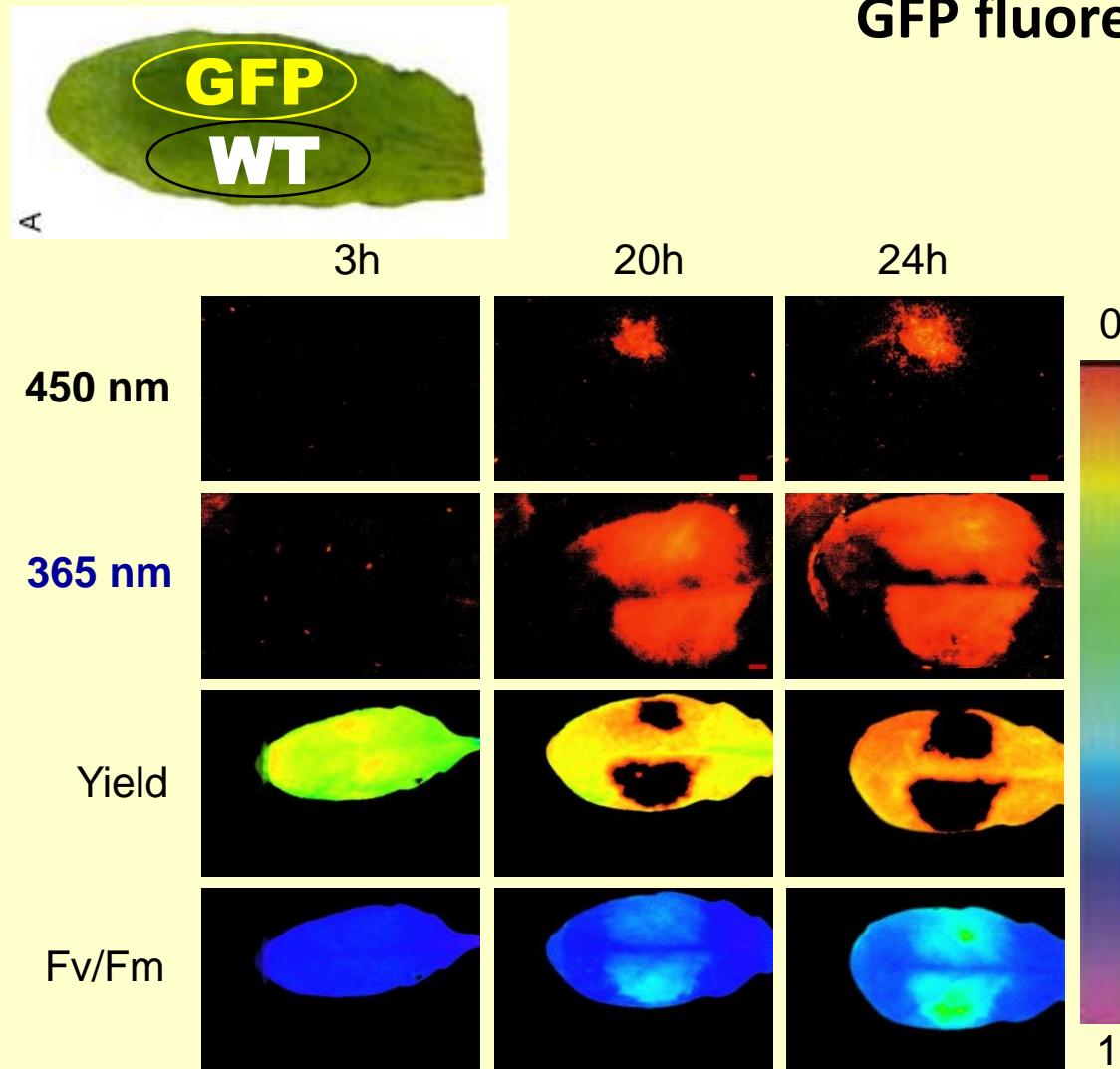
- Non invasive
- Very high sensitivity
- Spatial resolution

Combination of chlorophyll fluorescence imaging & statistical analysis: Combinatorial imaging



Development of multifluorescence imaging

Discrimination between chlorophyll, phenolic and GFP fluorescence



Simultaneous infiltration of WT strain and GFP labelled strain of *P. syringae*

PAM/GFP- measuring head

UV-light measuring head

PAM/GFP- measuring head

Enzyme activity signatures



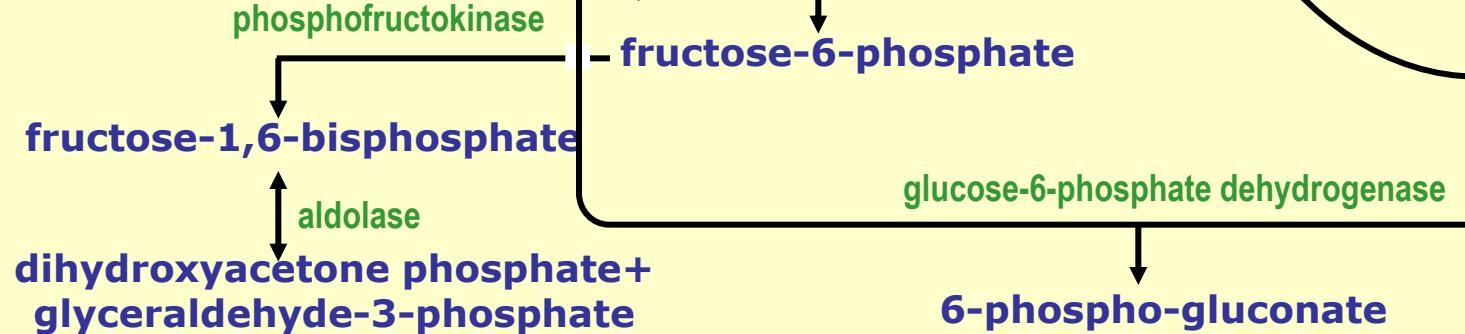
Uniform extraction :

- small sample size
- direct comparison

96-well format and

3 plate readers:

- semi-high throughput
- 300 assays/day
vs 30 in single cuvets



Characterisation of sugar beet development

Times course of enzyme activity signatures

Parameter [min/max]	No. 1	No. 2	No. 3	No. 4
Frischgewicht (0,01 – 25 g)	▲ ▲ ▲ ▲ ▲ ▲	▲▲▲ ▲▲▲	▲ ▲ ▲ ▲ ▲ ▲	▲ ▲ ▲ ▲ ▲ ▲
Suc-Gehalt (10 – 60.000 µg/gFW)	▲ ▲ x ▲ ▲ ?	▲▲ x ▲▲?	▲ ▲ x ▲ ▲ ?	▲▲ x ▲ ▲ ?
Sucrol. Aktiv. ●	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼
vacInv ●	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼
cytInv ●	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼
cwlInv ●	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼	▼▼▼▼▼▼
SuSy (nd – 2 nkat*mg ⁻¹ Prot) ●	▲ ▲ ▲ ▲ ▲	▲▲ — ▲▲▲	▲ — — ▲▲▲	▲ — — ▲▲▲
PFK (nd – 1 nkat*mg ⁻¹ Prot) ●	▲ ▲ — ▲▲▲	▲▲ — ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲
UGPase (nd – 6 nkat*mg ⁻¹ Prot) ●	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲
PGI (nd – 20 nkat*mg ⁻¹ Prot) ●	▲▲ ▲▲ ▲▲▲	▲▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲
PGM (nd – 20 nkat*mg ⁻¹ Prot) ●	▲▲ ▲▲ ▲▲▲	▲▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲
G6PDH (nd – 3 nkat*mg ⁻¹ Prot) ●	▲▲ ▲▲ ▲▲▲	▲▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲	▲▲ ▲▲ ▲▲▲
Aldolase (< 0,1 – 2 nkat*mg ⁻¹ Prot) ●	□ — □ □ □	□ — □□□□	□ □ □□□□	□ □ □□□□

▲ = Increase over time (90 days); ▼ = decrease over time; □ = transient;
 — = no change; x = n. d.; ? = no data yet

Instrumental analysis: Hormone levels

1. Cytokinins

a) biological active

- *trans*-Zeatin (tZ)
- *trans*-Zeatin-Ribosid (tZR)
- Dihydrozeatin (DHZ)
- Dihydrozeatin-Ribosid (DZR)
- Isopentenyl-Adenin (iP)
- *cis*-Zeatin (cZ)

b) conjugates:

- Isopentenyl-Adenin-7-Glucoside (IP7G)
- *trans*-Zeatin-O-Glucosid (ZOG)
- *trans*-Zeatin-O-Glucosid-Ribosid (ZOGR)
- *trans*-Zeatin-7-Glucoside (tZ7G)
- *trans*-Zeatin-9-Glucoside (tZ9G)

2) other hormones/ signal molecules

- Indolacetic Acid (IAA)
- Abscisic Acid (ABA)
- Jasmonic Acid (JA)
- Salicylic Acid (SA)

3) To be established: Gibberellins

- G A1
- G A3
- G A4
- G A5
- G A6
- G A8



UHPLC/Ms-Ms

Determination of spatial and temporal dynamics of plant responses during development within environmental constraints and interactions

